# ISOLATION OF RP 71955, A NEW ANTI-HIV-1 PEPTIDE SECONDARY METABOLITE 

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The genome of the human immuno deficiency virus (HIV-1) encodes a proteinase which processes the gag-pol and the gag polyproteins ${ }^{1)}$. Inhibition of these processing steps results in the production of progeny virions which are immature and non infectious ${ }^{2}$. Thus the HIV-1 protease, which belongs to the aspartic proteinase family ${ }^{3}$, has become an important target for the search of anti-HIV-1 compounds. In the course of our screening program to find inhibitors of this enzyme using a fluorometric assay ${ }^{4}$ ) we have isolated a novel metabolite.

For the production of this compound the Streptomyces strain SP9440 was inoculated into a $250-\mathrm{ml}$ Erlenmeyer flask containing 50 ml of seed medium consisting of corn steep $1 \%$, sucrose $3 \%$, calcium carbonate $7.5 \%$ and ammonium sulfate $2 \%$. The seed culture was incubated for 3 days at $28^{\circ} \mathrm{C}$ on a rotary shaker ( 250 rpm ). 5 to 12 ml of the culture was then transferred to a 2 -liter Erlenmeyer flask containing 250 ml of the same medium and the culture was incubated for 3 other days at $28^{\circ} \mathrm{C}$ on a rotary shaker ( 150 rpm ). The whole culture was transferred into a 100 -liter fermentor containing 45 liters of the same medium and incubated at $28^{\circ} \mathrm{C}$

Fig. 1. Structure of RP 71955.

under agitation ( 400 rpm ) and aeration ( $4 \mathrm{~m}^{3} /$ hour) for 2 days. 40 liters of the culture was then transferred into a 800 -liter fermentor containing 400 liters of a medium consisting of yeast extract $1 \%$, glucose $3 \%$, calcium carbonate $5 \%$, sodium chloride $2 \%$, potassium dihydrogen phosphate $0.1 \%$ and magnesium sulfate $0.1 \%$, and incubated at $28^{\circ} \mathrm{C}$ under agitation ( 150 rpm ) and aeration ( $15 \mathrm{~m}^{3} /$ hour) for 192 hours. The broth ( 440 liters) was centrifuged and the supernatant was discarded. The mycelium was extracted with acetone ( 300 liters) and then centrifuged. The pellet was discarded. The acetone was removed under reduced pressure from the supernatant and the resulting aqueous phase ( 60 liters) was applied on a Duolite S 861 stainless steel column ( $20 \times 60 \mathrm{~cm}$ ). The column was washed with water ( 60 liters) and eluted with $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}$ ( $1: 4$ ) ( 60 liters), $50 \% \mathrm{MeOH}$ ( 30 liters) and finally MeOH ( 60 liters) at a flow rate of $500 \mathrm{ml} /$ minute. The methanolic fraction which contains the active metabolite was concentrated under reduced pressure to yield an aqueous phase ( 10 liters) which was applied on a Diaion HP20 (Mitsubishi Chemical Industries Limited) glass column ( $10 \times 15 \mathrm{~cm}$ ) to adsorb the metabolite. The column was washed with water and eluted with MeOH ( 8 liters). This fraction was made $50 \% \mathrm{MeOH}$ and subjected to a preparative HPLC using an Amicon C18 $(20 \mu \mathrm{~m}$, 100 A ) stainless steel column ( $7.62 \times 50 \mathrm{~cm}$ ) fitted with a stainless steel precolumn. The column was first washed with $50 \% \mathrm{MeOH}$ ( 2 liters). A step gradient elution of increasing methanolic concentration from $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}(3: 2)$ to $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}(4: 1)$ was then performed and 800 ml fractions were collected at a flow rate of $200 \mathrm{ml} /$ minute. The active eluate ( $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}(7: 3$ ) and (4:1) fractions) was concentrated under reduced pressure to yield a crude powder $(9.2 \mathrm{~g})$. An aliquot ( 586 mg ) was dissolved in a mixture of MeOH and DMSO $10: 1$ and chromatographed on a Sephadex LH20 column ( $3 \times 150 \mathrm{~cm}$ ) developed with MeOH . Flow rate was $1 \mathrm{ml} /$ minute and 15 ml fractions were collected. The active fractions were collected and taken to dryness to yield 445 mg of a beige powder. An aliquot ( 223 mg ) was dissolved in 60 ml of $0.1 \mathrm{~N} \mathrm{HCl}-\mathrm{MeOH}$ (1.1) and applied on a Amicon C18 ( $20 \mu \mathrm{~m}, 60 \mathrm{~A}$ ) column $(2.5 \times 6 \mathrm{~cm})$. The column was washed with 60 ml of $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}(5.5: 4.5)$ and eluted with a step gradient of increasing methanolic concentration from $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}(6.5: 3.5)$ to $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}$ (9: 1). 5 ml fractions were collected at a flow rate of $10 \mathrm{ml} /$ minute. The active eluate ( $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}(4: 1)$,

Table 1. Physico-chemical properties of RP 71955.

| Appearance | White powder |
| :--- | :--- |
| $[\alpha]_{\mathrm{D}}^{20}$ | $-77^{\circ}(c 0.1, \mathrm{MeOH})$ |
| SI-MS $(\mathrm{M}+\mathrm{H})^{+}$ |  |
| Calcd for $\mathrm{C}_{9} \mathrm{H}_{132} \mathrm{O}_{26} \mathrm{~N}_{23} \mathrm{~S}_{4}$ | $2,164.4$ |
| Found: | $2,164.4$ |
| $\mathrm{UV} \lambda_{\text {max }}^{\mathrm{MeOH}} \mathrm{nm}(\varepsilon)$ | $283(6,200), 290(4,700)$ |
| IR $(\mathrm{KBr}) \mathrm{cm}^{-1}$ | $3400,3045,2945$, |
|  | $1740(\mathrm{sh}), 1670,1530$, |
|  | $1210,875,750,600$ |
| Rf value | 0.45 |
| Solubility |  |
| $\quad$ Soluble | MeOH, DMSO |
| Insoluble | Hexane, $\mathrm{CHCl}_{3}, \mathrm{EtOAc}$ |

a Merck Art. No. 5715, ethyl acetate-acetic acidwater (4:1.2:1).
( $8.5: 1.5$ ) and ( $9: 1$ ) fractions) was concentrated under reduced pressure to yield 143 mg of the pure metabolite as a colorless powder. The physicochemical properties of the metabolite are summarized in Table 1. The structure of RP 71955 was assigned by NMR studies ${ }^{5)}$ and was shown to be an original peptidic secondary metabolite having its $N$-terminal amino acid acylated by the $\beta$ carboxylic acid of the aspartic residue (Fig. 1). RP 71955 inhibited the HIV-1 aspartyl protease ( $\mathrm{IC}_{50}$, $35 \mu \mathrm{~g} / \mathrm{ml}$ ) and was found to be active in a T lymphocyte HIV-1 infection assay (A. Bousseau, personal communication) as measured by the inhibition of the cytopathogenic effect of the virus
( $\mathrm{IC}_{50}, 1$ to $3 \mu \mathrm{~g} / \mathrm{ml}$ ) as well as by the inhibition of the production of reverse transcriptase $\left(\mathrm{IC}_{50}\right.$, $4 \mu \mathrm{~g} / \mathrm{ml})$.

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